

TEICHOMYCINS, NEW ANTIBIOTICS FROM *ACTINOPLANES*
TEICHOMYCETICUS NOV. SP.

II. EXTRACTION AND CHEMICAL CHARACTERIZATION

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The teichomycins are new antibiotics produced by *Actinoplanes teichomyceticus* nov. sp. Teichomycin A₁ is a phosphorus-containing antibiotic, active *in vitro* and *in vivo* against gram-positive bacteria, and active only *in vitro* against gram-negative bacteria. Teichomycin A₂ is a chlorine-containing antibiotic, active *in vitro* and *in vivo* against gram-positive bacteria. Isolation, purification and physical and chemical properties of the two antibiotics are reported. Teichomycin A₁ is a member of the phosphoglycolipid class of antibiotics while teichomycin A₂ is related to the group of the glycopeptide antibiotics.

The teichomycins are a complex of antibiotics produced by *Actinoplanes teichomyceticus* nov. sp. ATCC N° 31121¹⁾. In this report the isolation and preliminary characterization of the two major components of the antibiotic mixture are described. The two products are referred to as teichomycin A₁ and teichomycin A₂.

Teichomycin A₁ is active *in vitro* against gram-positive and gram-negative bacteria and teichomycin A₂ is active *in vitro* only against gram-positive bacteria¹⁾. Both antibiotics inhibit bacterial growth by interfering with cell-wall synthesis²⁾. They are very effective in protecting mice which have been infected with medically important pathogens such as: *Staphylococci*, *Streptococci*, and *Diplococci*. They show low acute toxicity in mice¹⁾.

Isolation

Teichomycins produced during the fermentation of *Actinoplanes teichomyceticus*¹⁾ are present both in the filtered medium and in the mycelial cake and can be extracted from the fermentation broth by the procedure described in Fig. 1. The crude antibiotic preparation obtained from the mycelial cake and from filtrates were combined and purified by column chromatography on Sephadex LH-20, using *n*-propanol - ethyl acetate - 0.2 N NH₄OH (10: 7: 7) as eluent.

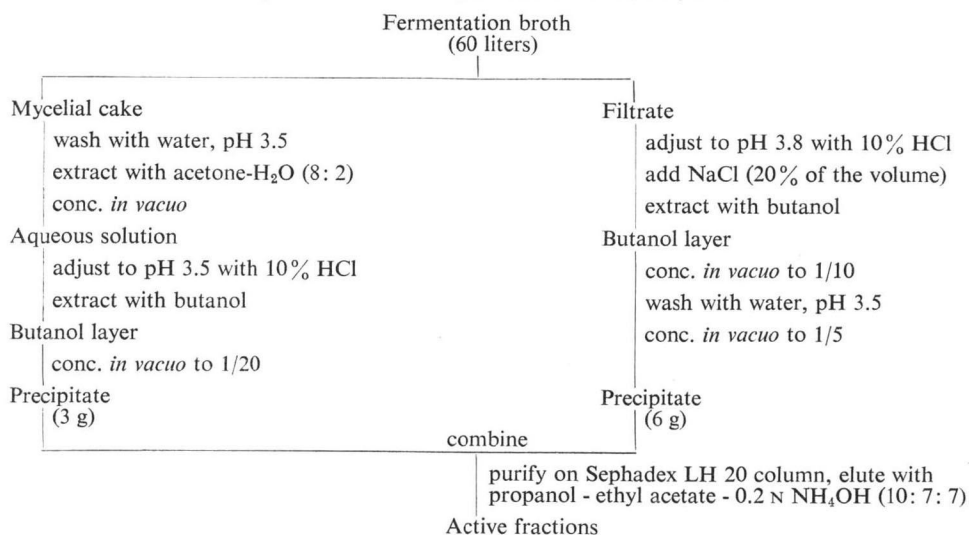
The fractions were analyzed by cellulose TLC using *n*-propanol - petroleum ether - conc.NH₄OH (4: 1: 4) as the solvent system with bioautography by *Staphylococcus aureus*. Teichomycin A₁ has an R_f 0.85 and teichomycin A₂ an R_f 0.6. The fractions corresponding to the two active products were collected and the antibiotics obtained as whitish amorphous powders.

Physical and Chemical Properties of Teichomycin A₁

Teichomycin A₁ was purified further by column chromatography on silica gel - Celite (1: 1) (v/v), using as eluent *n*-butanol - CH₃COOH - H₂O (8: 2: 2).

The antibiotic, which melts with decomposition at 220°C, is soluble in water at pH 7.0, in dimethylformamide and dimethylsulfoxide. It is partially soluble in methanol and ethanol, insoluble in

Fig. 1. Isolation and purification of teichomycins



diluted mineral acids and in non-polar organic solvents.

Teichomycin A₁ exhibits no characteristic UV-absorption between 220~360 nm. It gives positive FEHLING, TOLLENS, KMnO₄, anthrone and MOLISCH reactions and negative GRIESS and SCHIFF reactions. The IR spectrum is shown in Fig. 2. The elemental analysis, given as the average of four different analyses, is as follows: C 52.9, H 7.6, N 5.25, ash 3.26. Analyses of the ash revealed phosphorus. The presence of phosphorus in the molecule was confirmed by detection of radioactivity in a highly purified preparation of teichomycin A₁ produced in the presence of ³²P-phosphate. The percentage of P present in teichomycin A₁, as determined by chemical method, is 0.96.

The minimal molecular formula, tentatively determined for one atom of P in the molecule, is: C₁₄₂H₂₄₅N₁₂O₆₉P, with a calculated molecular weight of 3,255. Two different systems were used to determine the molecular weight of the antibiotic: analytical ultracentrifugation and gel filtration on Sephadex G-75. The molecular weight was established by ultracentrifugation on a Spinco Model E analytical ultracentrifuge and calculated with the SVEDBERG equation³¹, after the sedimentation and diffusion coefficients had been determined by the ultracentrifuge. The gel filtration on Sephadex G-75 was performed on a column 1.6×114 cm, with Blue Dextran and vitamin B₁₂ as internal markers.

The results are the following:

Solvent system	Ultracentrifuge	Sephadex G-75
Veronal buffer pH 8.5	17,700 daltons	
SÖRENSEN'S phosphate buffer pH 7.38		20,000 daltons
McILVAIN citrate buffer pH 4.4		30,000 daltons
9 M Formamide	57,400 daltons	

The antibiotic gave different molecular weights depending on the polarity and the pH of the solvent system used for the determination, suggesting that teichomycin A₁ aggregates in aqueous solutions. The size of the aggregates varied with the polarity and the pH of the solvent systems used for the determination. The lowest molecular weight value probably is that of a multimer, as the higher values are not regular multiples of the lowest one.

The behaviour of teichomycin A₁ in paper and thin-layer chromatography is shown in Table 1.

Table 1. Chromatographic behaviour of teichomycin A₁ and teichomycin A₂

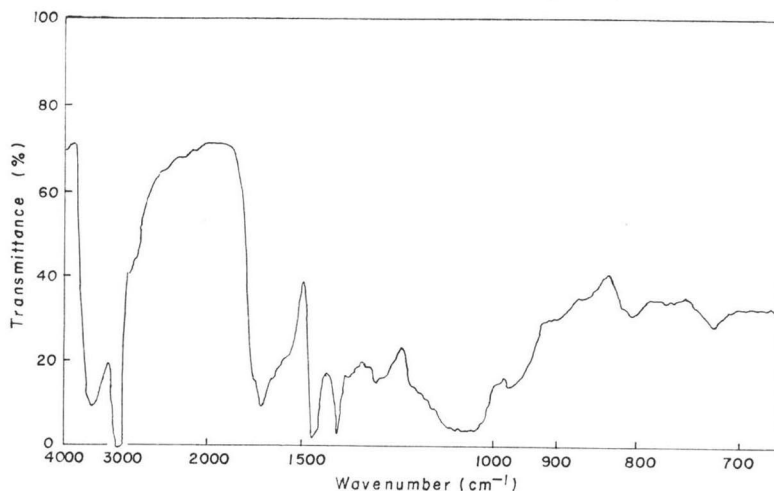
Elution systems	Rf values	
	Teichomycin A ₁	Teichomycin A ₂
°1) <i>n</i> -Butanol saturated with M/15 phosphate buffer, pH 6.0	0.0	0.0
°2) <i>n</i> -Butanol saturated with water containing 2% of <i>p</i> -toluenesulfonic acid	0.05	0.13
°3) <i>n</i> -Butanol saturated with water containing 2% ammonium hydroxide	0.0	0.0
°4) M/15 Phosphate buffer, pH 6.0, saturated with <i>n</i> -butanol	0.2	0.25
°5) 20% Aqueous solution of NH ₄ Cl	0.0	0.0
°6) <i>n</i> -Butanol - methanol - water (40: 10: 20) with 0.75% of methyl orange	0.42	0.37
°7) <i>n</i> -Butanol - methanol - water (40: 10: 20)	0.46	0.41
°8) Ethyl acetate saturated with water	0.0	0.0
°9) <i>n</i> -Propanol - <i>n</i> -butanol - 10 N NH ₄ OH (2: 3: 4)	0.55	0.43
*10) <i>n</i> -Propanol - ethyl acetate - conc. NH ₄ OH (2: 1: 2)	0.48	0.10
**11) <i>n</i> -Propanol - petroleum ether 35/60 - conc. NH ₄ OH (4: 1: 4)	0.85	0.6

° Paper chromatography on Whatman No. 1

* Thin-layer chromatography on silica gel.

** Thin-layer chromatography on cellulose.

The spots were visualized by microbiological assay with *Staphylococcus aureus*.

Fig. 2. Infrared spectrum of teichomycin A₁

Hydrolysis of the antibiotic in 2 N H₂SO₄ in a sealed tube at 100°C for 2 hours gave a mixture of at least three TOLLENS-positive products, two of which were also ninhydrin-positive. These reactions and the absorptions at 3300 and 1180 cm⁻¹ in the IR spectrum (Fig. 2) are indicative of the presence of carbohydrate residues in the molecule.

Furthermore, two other ninhydrin-positive products were found on the chromatographic analysis of the hydrolyzate of teichomycin A₁ (Fig. 3).

Physical and Chemical Properties of Teichomycin A₂

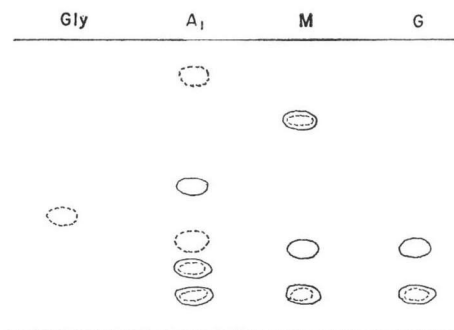
Teichomycin A₂ was dissolved in methanol - water (1: 1) and treated with acidic ion-exchange resin

(IR 120 or Dowex 50) up to pH 4.01 to remove the excess of base. After quick removal of the resin the antibiotic precipitated from the solution cooled at 4°C. Teichomycin A₂ is an amorphous powder which melts with decomposition at 260°C. The antibiotic is soluble in water at pH >7.0 and in methanol-water mixtures. It is partially soluble in methanol and in ethanol, insoluble in non-polar organic solvents. The antibiotic gives positive reactions with FEHLING, TOLLENS, KMnO₄, FeCl₃ and FOLIN-CIICALTEU reagents and negative reactions with GRIESS, anthrone and SCHIFF reagents. It gives a dark violet color with conc. H₂SO₄. The IR spectrum is shown in Fig. 4. The elemental analysis, given as the average of three different analyses is as follows: C 54.20, H 5.70, N 6.80, Cl 3.30. The minimum molecular formula, calculated for one atom of Cl in the molecule, is : C₄₈H₆₁N₅ClO₂₀, which corresponds to a minimum molecular weight of 1,062.

The UV spectrum shows an absorption maximum at 278 nm ($E_{1\text{cm}}^{1\%}$ 55) in acidic and neutral solutions and an absorption maximum at 297 nm ($E_{1\text{cm}}^{1\%}$ 74) in alkaline solution (Fig. 5). The behaviour of teichomycin A₂ at different pH's, with bathochromic shift from acidic to basic conditions, indicates the presence of ionizable functions related to the chromophore.

Potentiometric titrations with 0.1 N NaOH in methylcellosolve - H₂O (4:1) (v/v) gave a value of pK 7.4. The same titration made in the presence of 4% formaldehyde gave a pK 5.66. These values indicate the presence of a primary or secondary amino group which interacts with formaldehyde to

Fig. 3. Chromatographic pattern of teichomycin A₁ and moenomycin after acid hydrolysis



A₁ : Teichomycin A₁ after hydrolysis in 2 N HCl

Gly: Glycine

M : Moenomycin after hydrolysis in 2 N HCl

G : Glucose+Glucosamine

Paper chromatography on Whatman No. 1

Elution system: *n*-butanol - acetic acid - water (2:1:1)

Visualization systems: ninhydrin -----

TOLLENS ———

Fig. 4. Infrared spectrum of teichomycin A₂

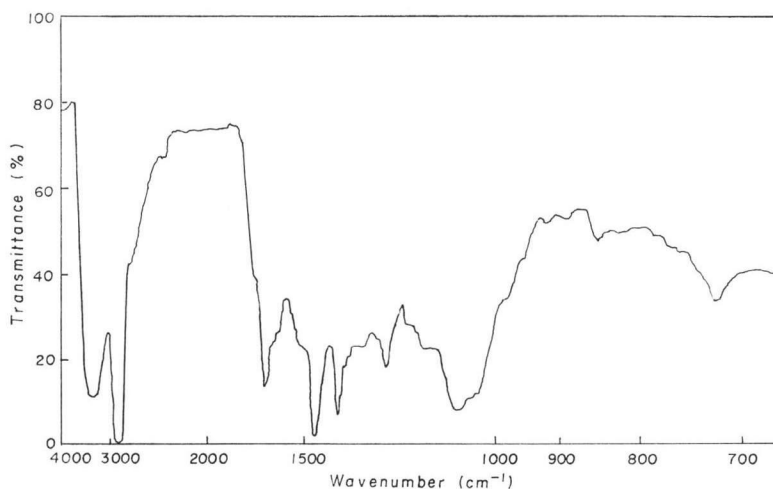
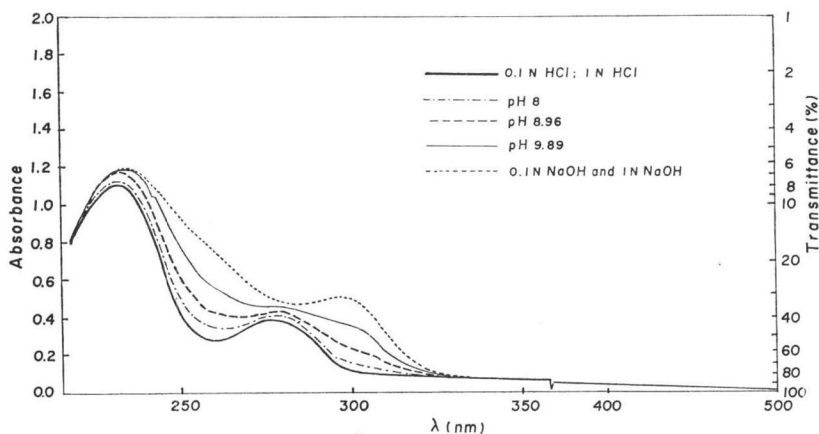


Fig. 5. Ultraviolet spectrum of teichomycin A₂

give a lower pK value. This basic function, titrated with NaOH, is present as a cation. Potentiometric titration with HClO₄ in dimethylsulfoxide solution indicated only one basic function. Potentiometric titrations of teichomycin A₂ with 0.1 N HCl in methylcellosolve - H₂O (4: 1) (v/v) gave a value of pK 4.95. This value must be related to an acidic group which, since it is titrated with acid, must be present as a salt.

The amphoteric character of the antibiotic has been confirmed by electrofocusing. The value p_I 6.5 was determined by thin-layer electrofocusing on polyacrylamide gel and microbiological assay with *S. aureus* as detecting system. The behaviour of teichomycin A₂ in paper and thin-layer chromatography is reported in Table 1.

Hydrolysis of the antibiotic in 2 N H₂SO₄ in a sealed tube at 100°C for 2 hours gave a mixture of two TOLLENS-positive products, one of which was also ninhydrin-positive. These reactions indicate the presence of two reducing sugars, including an aminosugar. These sugars have been isolated from the hydrolyzed mixture by means of ion-exchange resins. The acidic products obtained from hydrolysis were removed from the solution by adsorption on a basic ion-exchange resin (Amberlite IR 904). The neutral and basic products were adsorbed on an acidic ion-exchange resin (Dowex CG-50) and eluted with water and 0.1 N HCl. The TOLLENS-positive product was present in the neutral eluates and identified as D-mannose by comparison of its physico-chemical properties (Rf, [α]_D, IR, GC, MS) with the standard sample. The TOLLENS and ninhydrin-positive product was present in the acidic eluates and identified as D-glucosamine by comparison of its physico-chemical properties (Rf, [α]_D, IR, GC, MS) with the standard sample.

The acidic products adsorbed on the anionic resin were eluted with 0.1 N HCl; among them a substance was found which still showed some biological activity. It no longer contains sugar, indicating that loss of the D-mannose and D-glucosamine does not completely inactivate the teichomycin A₂ molecule. Hydrolysis in 6 N HCl sealed tube for 18 hours at 100°C gave a mixture of at least three ninhydrin-positive products, indicating the presence of aminoacids. The amide bands at 1660 cm⁻¹ (C=O_{st}) and at 1040~970 cm⁻¹ (C-O_{st}) in the IR spectrum of teichomycin A₂ are in accord with the presence of a peptide moiety.

Table 2. Chemical data on the glycolipid phosphorus-containing antibiotics

	P%	Formula	Glucose	Glucosamine	2NH ₂ -2, 6-dideoxyglucose	Other neutral carbohydrate	Glycine
Moenomycins	1.7~2	C ₇₀ H ₁₂₁ N ₅ O ₄₀ P	+	+	+	+*	n.r.
Diumycins	1.89~1.91	C _{68~72} H _{93~107} N ₅ O _{38~40} Na ₃ P	+	+	—	n.r.	—
Prasinomycins	2.43~2.28	C _{62~74} H _{96~130} N _{5~6} O _{32~40} P	n.r.	+	+	n.r.	+
Macarbomycins	2.12	C _{65~79} H _{123~144} N _{6~7} O _{41~48} P	+	+	—	n.r.	+
RP 11837	2.25	n.r.	n.r.	n.r.	n.r.	n.r.	n.r.
RP 8036	2.2	n.r.	n.r.	n.r.	n.r.	n.r.	n.r.
RP 19402	2.4	n.r.	n.r.	n.r.	n.r.	n.r.	n.r.
Teichomycin A ₁	0.96	C ₁₄₂ H ₂₄₅ N ₁₂ O ₆₉ P	—	+	—	+	—

* This pattern is related to an hydrolysis after 3~5 hours in 2 N HCl 100°C.

n.r. not reported

Discussion

The two antibiotics produced by *A. teichomyceticus* have different biological and chemical properties.

Teichomycin A₁ is characterized by the presence of phosphorus and the ability to form aggregates in solution, with the size depending on the pH and the polarity of the solvent. In contrast teichomycin A₂ contains no phosphorus, but does contain chlorine which is absent in teichomycin A₁. The properties described lead us to conclude that the two products are members of two different classes of antibiotics. The mechanism of action of teichomycin A₁ which inhibits cell-wall synthesis, the presence of phosphorus in the molecule and the property of aggregating in aqueous solvents suggest its similarity to the phosphoglycolipid antibiotics including the moenomycins⁴⁾, diumycins⁵⁾, prasinomycins⁶⁾, macarbomycins⁷⁾, RP 11837⁸⁾, RP 8036⁹⁾ and RP 19402¹⁰⁾. These antibiotics contain different neutral and basic sugars. The presence of neutral and basic carbohydrate residues was also established for teichomycin A₁. The chromatographic pattern of an acid hydrolyzate of teichomycin A₁ differed from the pattern for hydrolyzed moenomyacin as shown in Fig. 3. Further studies are in progress in order to identify the sugar moieties of teichomycin A₁. The data on teichomycin A₁, compared with those on the other phosphoglycolipid antibiotics are presented in Table 2. The percentage of phosphorus and the absence of glucose and of 6-deoxyglucosamine distinguish teichomycin A₁ from the other antibiotics of the class.

The mechanism of action of teichomycin A₂, its biological properties and the ability to give active products after hydrolysis suggest a similarity between this product and the glycopeptide class of antibiotics, such as vancomycin¹¹⁾, the ristomycins¹²⁾, the ristocetins¹³⁾, the actinoidins¹⁴⁾, LL-AV 290¹⁵⁾ and the mannopeptins¹⁶⁾.

Teichomycin A₂ differs from the known antibiotics of this class in that its net charge, is almost neutral (Pi=6.5) while it is basic for the others. Furthermore, teichomycin A₂ differs from the ristomycins, ristocetins, and mannopeptins in that teichomycin A₂ contains chlorine. Teichomycin A₂ can be distinguished from vancomycin by TLC analysis; teichomycin A₂, has an R_f 0.10 while vancomycin has an R_f 0.21 (solvent system 10 of Table 1).

Furthermore, teichomycin A₂ differs from the other glycopeptide antibiotics in carbohydrate composition as shown in Table 3. Teichomycin A₂ is the first antibiotic of this class which contains the basic carbohydrate D-glucosamine. In the other antibiotics, the basic carbohydrate is usually 2,5-dideoxy-2-amino-glucose or -arabinose. Mannose is present in 6 of 8 antibiotics, while glucose is

Table 3. Carbohydrate composition of the glycopeptide antibiotics

Antibiotic	Neutral carbohydrate	Basic carbohydrate
Vancomycin ¹⁸⁾	Glucose	Vancosamine
Actinoidins ¹⁹⁾	Glucose-mannose	Acosamine-actinosamine
Ristomycin A ²⁰⁾	D-Glucose, D-mannose, L-Rhamnose, D-arabinose	Ristosamine
Ristomycin B ²⁰⁾	D-Glucose, D-mannose, L-rhamnose	Ristosamine
Ristocetins ²¹⁾	Glucose, mannose, rhamnose, D-arabinose	Acosamine or ristosamine
LL-AV 290 ^{15,22)}	Glucose, rhamnose	Acosamine or ristosamine
Mannopeptins ¹⁶⁾	D-Mannose	No amino sugar
Teichomycin A ₂	D-Mannose	D-Glucosamine

present in every antibiotic except the mannopeptins and teichomycin A₂. Consequently we conclude that teichomycin A₂ is a new member of the glycopeptide class.

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